

MICHELSON PRIZE DATA PACKAGE-1 (DP-1) REQUIREMENTS

Applicants should refer to the **Michelson Prize and Grants Research Quality Assurance Toolkit** at www.michelsonprizeandgrants.org/resources/qa-toolkit for quality assurance practices and data that are required for Data Package-1. Toolkit references below refer to quality practices described on that website.

Section A. Preliminary Data

Preliminary data are defined as the entirety of results obtained in all studies completed from project inception up to the point immediately prior to the initiation of studies associated with the specific safety and effectiveness data sets required in sections B and C, respectively. The goal of this section is to provide a clear summary of the project history and scientific progress that led to the applicant’s decision to advance the program to complete the safety and effectiveness sections outlined below (sections B and C).

All *in vitro* and *in vivo* studies for which data sets exist must be presented and summarized in this section. Data presented cannot be pre-selected for, or only restricted to, positive safety and effectiveness results. It is important to include all negative results that may have failed to meet the safety and effectiveness target thresholds set for preliminary studies. Similarly, unexpected results, outcomes, and data that may be inconsistent with the original scientific theory and associated mechanism of action(s) hypothesis must also be presented.

Toolkit Section 5

This section must include a short summary for each of the following subsections:

1. Brief Description of Project History

- a. **Research Team, Institution(s), and Funding Source(s).** Provide a list of the research team members, showing the Principal Investigator, all Co-Investigators, and affiliated institutions(s) for each individual. The funding source(s) that supported all the preliminary work should be listed, including amount, duration, Principal Investigator, and funded study title(s).
- b. **Scientific Theory.** Provide a clear, succinct statement of the hypothesis supported through repeated testing results (e.g., accumulated evidence from preliminary data provided herein, or in published studies from other laboratories that support the principal hypothesis).

Toolkit Section 1

- 38 c. **Scientific Approach.** Provide a brief summary of scientific
 39 approach(es) used to test the hypothesis. Include description of the
 40 most important methods used to obtain all the relevant outcomes that
 41 were measured (e.g., clearly explain how safety and effectiveness were
 42 assessed).
- 43 d. **Animal Models Used.** Provide a summary of all laboratory animals
 44 (e.g., rodents, rabbits), cat, dog, and any other animal species used to
 45 generate the preliminary data.

Toolkit Section 3

46 2. Summary of Scientific Data – Provide all preliminary data for all *in vivo*
 47 (animal studies), *ex vivo*, and/or *in vitro* studies using samples from animal
 48 studies. For each study conducted, the following should be provided:

- 49 • **Study Title.**
- 50 • **Study Purpose.** State primary and secondary (if any) objectives.
- 51 • **Species and Sex.** List all laboratory animals, cat, dog, and any other
 52 animal species in which the candidate was tested. Indicate the
 53 gender (female and male) and reproductive status (pre-pubertal, post-
 54 pubertal).
- 55 • **Numbers Used.** List number of laboratory animals used.
- 56 • **Study Duration.** List the total length of the in-life laboratory animal
 57 phase, including any interim time points at which laboratory animals
 58 were euthanized.
- 59 • **Results (Outcomes) Measured.** Succinctly describe the techniques
 60 and methods used to measure each outcome measured.
- 61 • **Results.** Provide in the form of a table, figure, image, diagram, etc.
 62 Include separate legend and description for each data set presented in
 63 order for each result to stand on its own (without supporting
 64 discussion text).
- 65 • **Result Interpretation.** Provide a brief, interpretative summary for
 66 each result presented.

Toolkit Sections 3, 4

Toolkit Section 5

- 67 3. Summary of Scientific Results
- 68 • **Positive Outcomes** (successes). Briefly list all the results that
 69 support the scientific theory and/or satisfy (meet or exceed) the
 70 decision criteria for project progression to enter the Data Package-1
 71 preliminary safety and effectiveness phase (e.g., data presented in

Toolkit Section 5

72 sections B and C, below). Include all results that are consistent with
 73 results obtained from published studies from other laboratories using
 74 similar scientific approach(es) and readouts.

- 75 • **Negative Outcomes** (failures). List all the results that failed to
 76 support the scientific theory and/or did not meet study goals for
 77 safety and/or efficacy.
- 78 • **Unexpected Outcomes**. List all the results that were not predicted
 79 and/or influenced how the preliminary safety and effectiveness
 80 studies (presented below) were subsequently conducted. Include any
 81 results that differed from results obtained from published studies
 82 from other laboratories using a similar scientific approach(es) and
 83 readouts.

84

85 **Section B. Safety**

86 The purpose of this section is to provide a summary of all rodent and target species (cats
 87 and dogs) studies conducted under the minimum experimental designs outlined below.

88 Data sets are intended for use in defining the target dose for the DP-2 Pivotal Safety
 89 Study.

90 1. Dose Definition

91 a. **Target (1X) dose**. Provide the proposed dose (1X) for each species
 92 (rodent, cat, dog) and for each sex (and for reproductive status i.e. pre-
 93 or post-puberal?).

94 b. **Justification**. Provide a brief rationale and any relevant result
 95 summary information (e.g., preliminary data, section A) that support
 96 the 1X target dose selected for each species.

97 2. Toxicology Studies

98 Depending on the product candidate’s chemical or molecular structure,
 99 genotoxicity studies are required that should be consistent with the
 100 International Conference on Harmonization (ICH) Draft Guidance document
 101 [S2(R1) Genotoxicity Testing and Data Interpretation for Pharmaceuticals
 102 Intended for Human Use; 2008 – **Appendix 1**]. A standard test battery is
 103 suitable and must include a test for gene mutation in bacteria, and
 104 genotoxicity evaluation in mammalian cells *in vitro* (e.g., mouse lymphoma
 105 L5178Y *tk* gene mutation assay) and/or *in vivo* (micronuclei assay using

Toolkit Section 3

Toolkit Sections 3-5

106 rodent hematopoietic cells). Depending on the experimental design,
 107 integration of the *in vivo* genotoxicity assays into the dose escalation rodent
 108 safety study may be acceptable to the Foundation.

109 3. Rodent Safety Studies

110 These studies must be conducted and completed prior to initiation of the
 111 target animal safety studies. The purpose of these studies is to provide data
 112 generated under the minimum experimental design that is likely to be
 113 predictive of acceptable safety in the target animal species (Section B.4). In
 114 all studies, the product candidate must be administered one time by the
 115 identical route and method intended for the target animal safety studies.

Toolkit Sections 3-5

116 a. **Study Design (minimum)**. In order to evaluate both the long term safety
 117 and margin of safety of a treatment anticipated to have a permanent
 118 effect, the in-life phase must be at least 12 months in duration. At a
 119 minimum, the following 4 treatment groups must be included: placebo,
 120 1X (target) dose, a medium dose, and a high dose. The medium and high
 121 doses are defined by the applicant; however, the medium and high doses
 122 should be at least two and five times the 1X dose, respectively. Post-
 123 administration time points for data collection must include: acute (2-4
 124 weeks), intermediate (3 months), and final (12 months). Depending on
 125 the mechanism of action of the treatment, the Rodent Safety study may
 126 be designed as a combined safety and effectiveness study (see Rodent
 127 Effectiveness study, C1, below).

Toolkit Section 5

128 b. **Study Readouts**. At a minimum, the following five principal
 129 measurements of candidate safety must be conducted:

Toolkit Sections 3, 5

- 130 i. Daily observations. Must include general behavior and appearance.
- 131 ii. Weekly observations. Must include body weight, and brief physical
 132 examination.
- 133 iii. Gross pathology. Must include all major tissues/organs/body
 134 systems and reproductive tissues specified in **Appendix 2** at a
 135 minimum of the 3 required time points (acute, intermediate and final)
 136 in the study design.
- 137 iv. Histopathology. For all groups and time points, must include
 138 reproductive tissues, and, for high dose group, must include all major
 139 tissues/organs/body systems specified in **Appendix 2**. For medium

140 and low dose groups, if histopathology in high dose group identifies
 141 any pathology in a specific tissue/organ/body system, then that
 142 specific major tissue/organ/body system must also be analyzed at a
 143 minimum of the 3 required time points in the study design to define
 144 margin of safety.

145 v. Serum chemistry and hematology. Standard panels with minimum
 146 tests specified in **Appendix 3** must be run at a minimum prior to
 147 dosing and at the intermediate and final time points required in the
 148 study design.

149 c. **Study Results.** Copies of all raw data must be submitted. Appropriate
 150 statistical methods should be used to assess comparisons between
 151 placebo and treated animals in all measured parameters. Analyzed data
 152 for each of the five principal measurements of candidate safety should be
 153 presented in tables, figures, diagrams, images, or other summary formats.
 154 A separate legend and description for each analyzed data set should be
 155 included.

Toolkit Sections 3-5

156 d. **Study Discussion and Conclusions.** Interpretation of study results for
 157 each data set should be provided and main conclusions should be
 158 summarized.

159 e. **Safety Risk/Mitigation Strategy.** Safety knowledge gaps for which key
 160 information may be missing due to the experimental design selected
 161 and/or results obtained should be identified, together with any
 162 recommendations on how these gaps might be addressed through
 163 experimental design modifications in target animal safety studies.

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165 4. Target Animal (Cat and Dog) Safety Studies

Toolkit Sections 3-5

166 Target species safety studies must be conducted and completed as part of the
 167 Data Package-1 requirements. The purpose of these studies is to produce
 168 acute safety data using a study design that will determine the doses in the
 169 cats and dogs of each sex to be used in the Data Package-2 (DP-2) Pivotal
 170 Safety Study, to follow. Studies must be performed in pre- and post-puberal
 171 male and female dogs and cats, and the product candidate must be
 172 administered one time by the identical route, and method, intended for the
 173 DP-2 Pivotal Safety Study, to follow. Furthermore, the treatment should be

174 composed/formulated/manufactured in a way to assure similarity and
 175 consistency to the final form to be used after full development. All results
 176 (positive, negative, and unexpected) must be documented and presented.

177 a. **Study Design.** In-life phase must be at least 1 month in duration.

Toolkit Section 3

178 Treated animals of each species and sex should receive a single
 179 administration of the product candidate in the form and by the route
 180 expected to be used in the later DP-2 pivotal safety study. The dose is
 181 defined by the applicant, but, at a minimum, must be at least twice the
 182 target (1X) dose. The principal aim is to identify any treatment adverse
 183 effects (AEs), generally defined as any unexpected side effects or
 184 unintended changes in the structure, function, or chemistry of the body,
 185 including injury, toxicity, and sensitivity reactions.

186 b. **Study Readouts.** At a minimum, the following four categories of
 187 observations of candidate safety must be included:

Toolkit Sections 3-5

- 188 i. Daily observations. Must include general behavior, body
 189 appearance, and food consumption.
- 190 ii. Weekly observations. Must include physical examination, body
 191 weight, standard serum chemistry and hematology profiles
 192 (**Appendix 3**), and standard urinalysis (**Appendix 4**).
- 193 iii. Other. Depending on mechanism of action, other readouts may be
 194 required, such as electrocardiogram data, bone density measurement,
 195 etc.
- 196 iv. Gross pathology and histopathology (**Appendix 2**). Euthanasia is
 197 not required in this study, but, if any animals die, a full necropsy is
 198 required, with complete histopathology performed to determine, if
 199 possible, the cause of death.

200 c. **Study Results.** Copies of all raw data must be included. Appropriate
 201 statistical methods must be used to assess comparisons between placebo
 202 and treated animals in all measured parameters. Analyzed data for all
 203 daily and weekly observation readouts of candidate acute safety should
 204 be presented in tables, figures, images, or other summary formats. A
 205 separate legend and description for each analyzed data set should be
 206 included.

Toolkit Section 5

- 207 d. **Study Discussion and Conclusions.** Interpretation of study results for
 208 each data set should be provided and main conclusions should be
 209 summarized.
- 210 e. **Safety Risk/Mitigation Strategy.** Safety knowledge gaps for which key
 211 information may be missing due to the experimental design selected
 212 and/or results obtained should be identified, together with any
 213 recommendation on how these gaps might be addressed through
 214 experimental design modifications in the DP-2 Pivotal Safety Study.
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216 **Section C. Effectiveness**

217 These studies must be conducted and completed for Data Package-1. The goal of this
 218 section is to provide a comprehensive summary of rodent and target species (cats and
 219 dogs) studies used to define the target effectiveness dose for later use in the DP-2 non-
 220 pivotal target host effectiveness study that will follow Foundation acceptance of DP-1.
 221 For each effectiveness study outline below, clearly stated definitions must be provided
 222 for:

- 223 • “Suppression of fertility and ablation of sex steroids and/or their effects”
 224 using at least one scientifically accepted or scientifically valid measurement.
- 225 • “Probability of permanence” using at least one scientifically accepted or
 226 scientifically valid measurement OR valid justification based on the product
 227 candidate mechanism(s) of action.

228 In all studies, the product candidate must be composed/formulated/manufactured in a way
 229 to assure similarity and consistency to the final composition intended for DP-2 non-
 230 pivotal target host efficacy study and during full development. The product candidate
 231 must be a single administration by the identical route and delivery method intended for
 232 the DP-2 non-pivotal target host efficacy study and during full development. Post-
 233 puberal animals should be used unless the applicant can justify that the effect will be
 234 different in pre-pubertal animals. All results (positive, negative, and unexpected) must be
 235 documented and presented.

236 1. Rodent Studies

- 237 • Must demonstrate complete suppression of fertility and ablation of sex
 238 steroids and/or their effects in both females and males for a minimum of
 239 12 consecutive months.

Toolkit Sections 3-5

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- Must demonstrate probability of permanence through data trend line that shows no expectation of recovery or rebound.
 - May need to demonstrate longer than 12 consecutive months depending on mechanism of action or if data trend line shows possible recovery or rebound.
 - Depending on the mechanism of action of the treatment, the Rodent Effectiveness Safety study may be designed as a combined safety and effectiveness study (see Rodent Safety study, B3, above).
2. Female Dog Studies
- Because the inter-estrous interval in female dogs is variable and can be up to 24 months, this study must demonstrate complete suppression of fertility (no estrous cycle) and ablation of sex steroids and/or their effects for a minimum of 24 consecutive months.
 - Must demonstrate probability of permanence through data trend line that shows no expectation of recovery or rebound.
 - May need to demonstrate effect for longer than 24 consecutive months depending on mechanism of action or if data trend line shows possible recovery or rebound.
3. Male Dog Studies
- Must demonstrate complete suppression of fertility and ablation of sex steroids and/or their effects for a minimum of 12 consecutive months.
 - Must demonstrate probability of permanence through data trend line that shows no expectation of recovery or rebound.
 - May need to demonstrate effect for longer than 12 consecutive months depending on mechanism of action or if data trend line shows possible recovery or rebound.
4. Male and Female Cat Studies
- Must demonstrate suppression of male and female fertility and ablation of sex steroids and/or their effects for a minimum of 12 consecutive months in cats housed under constant or long (>14 hours) day-length photoperiod.
 - Must demonstrate probability of permanence through data trend line that shows no expectation of recovery or rebound.

- 273 • May need to demonstrate effect for longer than 12 consecutive months
 274 depending on mechanism of action or if data trend line shows possible
 275 recovery or rebound.

276

277 **Section D. Mechanism of Action**

278 The primary objective of this section is to define the mechanism(s) by which the
 279 pharmacologically active substance (e.g., drug or vaccine) or device generates the
 280 antifertility (e.g., sterilant) effect in rodents, cats and dogs. Provide answers or address
 281 each of the following:

Toolkit Sections 3, 5

- 282 1. How does the product candidate work? Identify and describe the specific target
 283 and/or biochemical pathway by which the candidate exerts its mode of action
 284 (MOA) (sterilant effect). If the product candidate has more than one MOA, each
 285 target should be discussed separately, followed by a brief summary of any known
 286 additive or synergistic anti-fertility effects.
- 287 2. If applicable, define the structure-activity relationship (SAR). Describe the
 288 relationship of the chemical or molecular structure of the product candidate with
 289 respect to its biological activity. If the structure has been quantitatively
 290 correlated with the biological activity (QSAR), provide the mathematical
 291 relationship (formula) used to predict the biological response of related chemical
 292 structures.
- 293 3. Why does the product candidate result in permanent suppression of fertility and
 294 ablation of sex steroids and/or their effects? List the measurable scientific
 295 parameters on which the conclusion is based for each species. Examples include
 296 (but are not limited to): serum testosterone, serum estradiol, serum progesterone,
 297 and testicular and ovarian histology.
- 298 4. Based on the MOA, provide any safety related issues or concerns that will be
 299 important to monitor in the DP-2 pivotal safety study, including need for
 300 personal protective equipment for persons handling the product candidate.

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302 **Section E. Pre-Final Formulation Composition**

Toolkit Sections 3, 5

303 If the treatment is a drug, this information is required. This section provides a foundation
 304 for the Chemistry, Manufacturing, and Controls (CMC) technical section of the FDA
 305 Center for Veterinary Medicine New Animal Drug Application (NADA). The objective

306 of this section is to provide assurance that the product can be made at commercial scale in
307 a manner that would be acceptable for approval by the FDA.

308

309 Provide a comprehensive list and description of all the individual components in the
310 product candidate. List each ingredient that will be used to make the product.

- 311 1. Outline how the product candidate might be made at commercial scale.
312 Demonstrate that there is reasonable probability that the compound can be
313 made at commercial scale by, for example, describing the key steps in the
314 outline of production and manufacture scale at which the product is most
315 likely to be produced.
- 316 2. Provide a concept of labeling. Describe the proposed immediate container
317 and outer packaging. Describe the type of vial, bottle, syringe, and/or packet
318 that the product itself will be in contact with.
- 319 3. Storage conditions and shelf life. Provide a summary of any real-time or
320 accelerated stability studies conducted to evaluate degradation, by-products,
321 solubility, moisture content, etc.

322

323 **Section F. Statistical Justification of Sample Size and Statistical Analysis**

Toolkit Sections 3, 5

324 The objective of this section is to provide a statistical justification of the number of
325 animals used in each treatment group and in each study completed to assess product
326 candidate safety and effectiveness. Describe the statistical methods and power analysis
327 used to determine treatment sizes to analyze the safety and effectiveness results for each
328 of the safety and effectiveness studies listed above.

329

330 **Section G. Quality Assurance**

331 The purpose of this section is to provide a summary of the quality, scientific practices
332 used to assure the reliability, reproducibility, and relevance of the entirety of scientific
333 data provided in Data Package-1.

334

335 The Michelson Prize & Grants program administrators recognize that all grant applicants,
336 grantees, and prize registrants are committed to performing sound scientific research with
337 traceable and repeatable processes that assure data integrity and confidence in research
338 findings. It is understood that studies in Data Package-1 may not be conducted under
339 GLP (Good Laboratory Practices) and GCP (Good Clinical Practices), as these studies

340 are not intended to be pivotal for regulatory approval. Therefore, in order to support the
341 recommended voluntary quality practices of researchers funded by FAF, the Foundation
342 has provided a **Quality Assurance Toolkit** to help researchers meet quality assurance
343 objectives as a means to promote the credibility and acceptability of their work. The FAF
344 Quality Assurance Toolkit was designed to provide opportunities for incorporating
345 quality assurance features into typical research environments and was based on selected
346 recommendations provided by the World Health Organization (WHO Handbook: Quality
347 practices in basic biomedical research, ISBN; 92 4 159445 4, 2006) and the British
348 Association for Research Quality Assurance (BARQA, Guidelines for Quality in Non-
349 Regulated Scientific Research, ISBN: 1-904610-08-0, 2006).

350

351 Use of the FAF Quality Assurance Toolkit will provide assurance that the quality critical
352 research practices associated with personnel and equipment management and the
353 recommended control of procedures, data, records, and methodology have been
354 implemented throughout the research investigation. References to toolkit sections that
355 will support DP-1 requirements are provided within this document.

356

357 It is recommended that all studies conducted adhere to general quality assurance
358 expectations and guidelines as demonstrated by a commitment to an established quality
359 standard, the implementation of a laboratory quality assurance management system, or
360 the use of the FAF Quality Assurance Toolkit in order to sharpen the focus on the sound
361 science and quality practices present in the research program. A site visit to evaluate
362 laboratory capacity to have generated DP-1 data and use, if any, of Toolkit items, will be
363 scheduled upon submission of DP-1 in order to review quality management practices.

364

365 **Section H. Supporting Information**

366 The objective of this section is to include all information about the product candidate that
367 was not provided in the other sections. Examples of information for this section include:

368

- 369 1. Project publications. Provide copies of all peer-reviewed publications,
370 publications in press, accepted for publication, or submitted manuscripts
371 associated with any of the data provided in Data Package-1.

371

- 372 2. Project summary reports. Provide copies of all draft manuscripts and all reports
generated to meet any funding requirements.

- 373 3. Relevant references. Provide list of key references that support the scientific
374 theory, approach, animal models, and results associated with any of the data
375 provided in Data Package-1.

376

APPENDICES

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378

Appendix 1. International Conference on Harmonization (ICH) Guidance

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Document [S2(R1) Genotoxicity Testing and Data Interpretation for

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Pharmaceuticals Intended for Human Use; 2012.]

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Appendix 2. Major tissues and organs to be collected for gross and

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histopathology in Study Readouts, minimum parameters.

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Appendix 3. Serum chemistry and hematology panels, minimum

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parameters.

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Appendix 4. Urinalysis, minimum parameters.

389 **Appendix 1.** [INTERNATIONAL CONFERENCE ON HARMONIZATION \(ICH\) GUIDANCE](#)
390 [DOCUMENT \[S2\(R1\) GENOTOXOCITY TESTING AND DATA](#)
391 [INTERPRETATION FOR PHARMACEUTICALS INTENDED FOR HUMAN](#)
392 [USE; 2012\].](#)

393 **Appendix 2. MAJOR TISSUES AND ORGANS TO BE COLLECTED FOR GROSS AND**
 394 **HISTOPATHOLOGY IN STUDY READOUTS, MINIMUM PARAMETERS.**

Tissue	Organ Weight Taken	Collected and Preserved	Microscopic Examination
Adrenal gland	X	X	X
Aorta		X	X
Bone with bone marrow, femur		X	X
Bone with bone marrow, sternum		X	X
Bone marrow smear ^a		X	
Brain (cerebrum, midbrain, cerebellum, medulla/pons)	X	X	X
Epididymis	X	X	X
Esophagus		X	X
Eye (with optic nerve)		X	X
Heart	X	X	X
Kidney	X	X	X
Large intestine, cecum		X	X
Large intestine, colon		X	X
Large intestine, rectum		X	X
Liver	X	X	X
Lung with bronchi		X	X
Lymph node, mandibular		X	X
Lymph node, mesenteric		X	X
Mammary gland (process females only)		X	X
Nerve, sciatic		X	X
Ovary (record morphology, presence of follicles and corpora lutea)	X	X	X
Oviducts		X	X
Pancreas		X	X
Peyer's patch		X	X
Pituitary	X	X	X
Prostate		X	X
Salivary gland, mandibular ^b	X	X	X
Salivary gland, parotid		X	X
Salivary gland, sublingual		X	X
Seminal vesicles with coagulating gland	X	X	X
Skeletal muscle, biceps femoris		X	X
Skin		X	X
Small intestine, duodenum		X	X
Small intestine, ileum		X	X
Small intestine, jejunum		X	X
Spinal cord, cervical		X	X
Spinal cord, lumbar		X	X
Spinal cord, thoracic		X	X
Spleen	X	X	X
Stomach, glandular		X	X

Tissue	Organ Weight Taken	Collected and Preserved	Microscopic Examination
Stomach, nonglandular		X	X
Target Organs ^c		X	X
Testis	X	X	X
Thymus	X	X	X
Thyroid gland (with parathyroid) ^d	X	X	X
Trachea		X	X
Urinary bladder		X	X
Uterus with cervix	X	X	X
Vagina		X	X
Gross lesions		X	X
Tissue masses with regional lymph node ^e		X	X

395 ^a Bone marrow smears will be prepared only for animals necropsied at scheduled intervals. Evaluation will be performed at the
396 discretion of the Study Director and/or Sponsor.

397 ^b The combined weight of the right mandibular/sublingual salivary gland will be obtained.

398 ^c Target organs (and target organ gross lesions) will be designated by the Study Director, Pathologist and/or Sponsor based on
399 experimental findings.

400 ^d Parathyroids cannot always be identified macroscopically. They will be examined if in the plane of section and in all cases
401 where they are noted as grossly enlarged.

402 ^e A regional lymph node drains the region where a tissue mass is located. A regional lymph node may not always be identified
403 when a mass is present.

404 **Appendix 3. SERUM CHEMISTRY AND HEMATOLOGY PANELS, MINIMUM PARAMETERS.**405 **Required Serum Chemistry Panel**

- 406 • alkaline phosphatase
- 407 • total bilirubin (with direct bilirubin if total bilirubin exceeds 1
- 408 mg/dL)
- 409 • aspartate aminotransferase
- 410 • alanine aminotransferase
- 411 • gamma glutamyl transferase
- 412 • sorbitol dehydrogenase
- 413 • urea nitrogen
- 414 • creatinine
- 415 • total protein
- 416 • albumin
- 417 • globulin and A/G (albumin/globulin) ratio (calculated)
- 418 • glucose
- 419 • total cholesterol
- 420 • triglycerides
- 421 • electrolytes (sodium, potassium, chloride)
- 422 • calcium
- 423 • phosphorus
- 424 • urea

425 **Required Hematology Panel**

- 426 • leukocyte count (total, relative and absolute differential)
- 427 • erythrocyte count
- 428 • hemoglobin
- 429 • hematocrit
- 430 • mean corpuscular hemoglobin
- 431 • mean corpuscular volume
- 432 • mean corpuscular hemoglobin concentration (calculated)
- 433 • absolute reticulocytes
- 434 • platelet count
- 435 • blood cell morphology
- 436 • prothrombin time
- 437 • activated partial thromboplastin time
- 438

439 Appendix 4. URINALYSIS, MINIMUM PARAMETERS.

- 440 • volume
- 441 • specific gravity
- 442 • pH
- 443 • color and appearance
- 444 • protein
- 445 • glucose
- 446 • bilirubin
- 447 • ketones
- 448 • blood
- 449 • urobilinogen
- 450 • microscopy of centrifuged sediment